

## The role of the ketogenic diet in exercise performance

Evan E. Schick

*Physiology of Exercise Sport (PEXS) Laboratory, California State University, Long Beach, California, 90840*

**Abstract.** Muscle glycogen storage and degradation are nearly universally accepted as crucial metabolic processes for ensuring adequate intramuscular energy levels during prolonged, high-intensity activity. However, a growing body of data illustrates that alternative substrates, such as ketone bodies, may be equally as effective in transducing energy during exercise. Ketosis, wherein ketones serve as the primary oxidative fuel, can be achieved nutritionally through a low-carbohydrate, high-fat ketogenic diet (KD). Though the KD is unequivocally successful in facilitating weight-loss with minimal sacrifice to lean mass, current research indicates a complex role for the KD in both anaerobic and aerobic exercise performance. This review discusses: 1) the mechanisms behind KD adaptation, and the effect of KD adaptation on 2) glycogen metabolism, 3) aerobic exercise performance and 4) anaerobic exercise performance.

**Key words:** *low-carbohydrate, high-fat, metabolism, athlete.*

### Introduction

Use of a low-carbohydrate, high-fat ketogenic diet (KD) is widely recognized as an efficacious therapy for a range of metabolic and neurodegenerative diseases and cancers (1-7). Clinical use of the KD has gained popularity as its effects often mirror those reached pharmacologically yet are attained with little off-target risk). However, interest in the KD extends beyond the clinical landscape as mounting evidence suggests that the KD may also influence exercise performance and adaptation). Though its precise role within exercise training and performance remains elusive, the notion that the KD might enhance exercise performance remains contentious, as it challenges traditional, carbohydrate (CHO)-centric guidelines for exercise and sport nutrition. Though past reviews have cohesively and comprehensively summarized ketone biochemistry and its clinical role, this paper aims at unifying current findings most directly implicated in exercise performance including: 1) the cellular mechanisms of KD adaptation as well as (2) the effects of KD on glycogen metabolism, 3) aerobic exercise and 4) anaerobic exercise (8-12).

Due to its storage abundance, intracellular location and rapid energy provision, muscle glycogen has long been held as the most important energy substrate during prolonged, high intensity exercise. This belief has resulted in the decades-long practice of CHO loading prior to competition among endurance athletes looking to achieve supra-maximal glycogen levels (13). The high fat and low CHO nutrient apportionment of the KD conflicts with conventional, CHO-centered sports nutrition guidelines, which recommend up to 12 grams/kilogram body weight for those engaged in high intensity endurance programs (14). Depending on the goal of the individual, the KD may be hypocaloric, eucaloric or hypercaloric and is typically comprised of a 3:1 to 4:1 energy ratio of fat to protein and CHO; though CHO restrictive diets with lower ratios of fat to protein can also be ketogenic (2, 15).

The biochemical underpinning of the KD is hepatic synthesis of ketone bodies, Acetoacetate (AcAc) and H- $\beta$ -hydroxybutyrate ( $\beta$ HB). Hepatic ketogenesis, upregulated as a consequence of limited carbohydrate supply and abundant fatty acid availability, converts acetyl CoA's, generated at rates prohibiting tri-carboxylic acid cycle entry, to AcAc and  $\beta$ HB (16). Both AcAc and  $\beta$ HB are transported via systemic circulation to extrahepatic tissues, where they are oxidized as needed (16, 17). In healthy fed adult humans, ketone oxidation represents only a minor fraction of total body energy expenditure, however, its contribution to energy metabolism in the heart, brain and muscle significantly increases in many physiological and pathological states including the neonatal period, fasting, starvation, repressed insulin production, insulin resistance prolonged exercise and low-carbohydrate diets (2, 11, 12, 17-19).

In fact, ketones -mainly  $\beta$ HB- supply up to 70% of the energy used by the brain during starvation with the remainder provided by endogenously derived glucose (17). Traditionally, in the before mentioned conditions, ketosis is seen as a metabolic means of holding serve until an adequate supply of blood glucose is available.

### **Mechanisms of KD Adaptation**

Understanding the mechanisms by which the body responds to the considerable shifts in macronutrient consumption and subsequent fuel utilization is crucial, as adaptation to a KD must be attained in order to normalize or improve performance. Dietary fat, ingested in substantial quantities to induce a state of ketosis, becomes the primary oxidative fuel by mass action. Subsequently, active tissues undergo a two-fold response to accommodate elevated fatty acid flux: increased mitochondrial  $\beta$ -oxidation and reduced glucose oxidation. Ample support for this tissue-level response has been provided, however, it is imperative to understand the cellular mechanisms responsible for this metabolic shift. Heightened  $\beta$ -oxidation is accomplished through an adaptive cellular response observed in studies of both animals and trained cyclists wherein prolonged KD (>1 week) amplified the activity of skeletal muscle carnitine acyltransferase (CAT) and hydroxyacyl-coenzyme A dehydrogenase (3-HAD) relative to the activity of citrate synthase (CS) (20-24). Analogous gene expression data has revealed a pointed prioritization of fat oxidation over its storage as upregulation of CD36, butyrate dehydrogenase (HBDH) and mitochondrial uncoupling protein-2 (UCP-2) was concomitant to reduced expression of fatty acid synthase (FAS) in the livers of KD-adapted mice (25). Similar studies of high-fat diet adaptation provide evidence of depressed glucose oxidation associated with reductions in pyruvate dehydrogenase activity and both insulin and exercise stimulated muscle glucose transport (26, 27). Provided that endurance training boosts capacity for fat oxidation, these tissue and cell-level adaptations suggest that the KD might catalyze physiological responsiveness to an endurance exercise program (28).

### **KD Adaptation and Glycogen Utilization**

Significant evidence links adaptation to the KD to altered muscle glycogen metabolism. Short term (3-14 days) KD has been reported to decrease baseline liver and muscle glycogen levels in rats and trained cyclists (20, 29, 30). However, in a study of moderately obese subjects following six weeks of a KD, resting muscle glycogen content was 57% of baseline after week 1 but increased to 69% of baseline values after week 6 (31). Newer findings demonstrate that ultra-endurance runners fully habituated to the KD (>20 months) experience similar baseline and post-exercise muscle glycogen levels compared to controls on a mixed-diet (32). These studies indicate that initial depletion of muscle glycogen induced by the KD may be at least partially reversed through habituation to the diet. Moreover, normal post-exercise glycogen repletion in KD-adapted individuals may be attained by increased lactate-mediated hepatic glucose production i.e. gluconeogenesis, and muscle glycogen synthase activity, especially in type II fibers (26, 33-35). Additionally, KD-adaptation has been shown to preserve liver and muscle glycogen during exercise; KD-adapted rats and trained humans have been shown to exhibit reduced exercising muscle and liver glycogen degradation rates without sacrifice to endurance performance (20-22, 29, 35, 36). These evidence, though somewhat indirect, suggest that chronic reliance on fat as a primary fuel increases its inherent rate of oxidation, an especially useful adaptation that reduces muscle glycogen degradation while maintaining cell energy levels during exercise (36).

### **Ketogenic Diet in Practice**

#### *Ketogenic Diet and Weight Management*

Perhaps the most understood and implemented application of the KD is within bariatric medicine, as obese individuals undertaking a hypocaloric KD experience a significant reduction in weight that is vastly attributed to fat loss with minimal loss in muscle mass (18, 37-39). Less studied, yet perhaps equally intriguing, is apparently even a *eucaloric* KD may promote fat loss and lean mass preservation in both obese and healthy weight individuals (15, 40, 41). Most recently, investigators reported that obese and overweight individuals following either an eight-week resistance training (RT) with a eucaloric, carbohydrate restrictive diet (<30grams/day) or RT with a conventional, hypocaloric diet saw equal improvements in fat composition and strength (42). Driving these KD-induced anthropometric adaptations is increased sensitivity to both insulin and thyroid hormone, which helps to limit rates of skeletal muscle catabolism during periods of weight-loss (15, 43).

These findings lend intriguing sports nutrition research opportunities for weight-category athletes who must reduce body mass while retaining muscle mass during competition preparation (43).

The KD could represent a healthier nutritional strategy for these athletes compared to commonplace rapid weight loss practices such as severe caloric and hydration restriction and induction of hyperthermia.

#### *Ketogenic Diet and Aerobic Exercise*

The manner in which KD affects aerobic performance appears to be determined by three factors: exercise intensity, training status and length of diet habituation. Bergstrom et. al., 1967, (12) illustrated that three days of a KD was enough to compromise submaximal endurance (75%  $\text{VO}_2\text{max}$ ) in healthy, untrained individuals. Similarly, a six-week high fat diet (HFD) significantly reduced work output in untrained subjects during a 45-minute bicycle test. The authors attributed this finding to an increased proportion of fat oxidation observed through decreased exercising respiratory exchange ratio (RER) (40). However, the purported sacrifice to high intensity exercise performance may represent a tradeoff for increased fatigue resistance. Indeed, moderately obese subjects adhering to a six-week KD exhibited reduced exercise intensity but greater stamina during a treadmill exercise test to subjective exhaustion (31). These studies suggest that enhanced fat oxidation may not be energetically compatible with high intensity exercise but it may augment long duration exercise.

Interestingly, aerobic performance in KD-adapted endurance athletes appears to be less susceptible to shifts in energy substrate utilization. Initial studies in trained cyclists revealed that 4-weeks of a KD did not affect moderate intensity (~65%  $\text{VO}_2\text{max}$ ) endurance exercise performance; apparently, enhanced exercising fat oxidation rates, observed through lower RER, were able to compensate for reductions in steady state glucose oxidation rates (36). More recently, elite ultra-endurance runners habituated (>20 months) to a low-carbohydrate diet (LC) displayed significantly greater rates of fat oxidation and lower rates of carbohydrate oxidation during a 180-minute run at 64% of  $\text{VO}_2\text{max}$ . Peak fat oxidation rates were also significantly greater in the LC group and were reached at a higher percentage of  $\text{VO}_2\text{max}$  compared to controls (32). Furthermore, following just two-weeks of a high fat diet (~70% fat), trained cyclists experienced lower RER values and enhanced endurance during moderate intensity (60%  $\text{VO}_2\text{max}$ ) exercise (29). More notably, HFD did not impair performance during a high-intensity (85%  $\text{VO}_2\text{max}$ ) time to exhaustion test. Likewise, 6-15 days of HFD enhanced fat oxidation and decreased CHO oxidation rates in trained cyclists while either not affecting or even improving performance during 20-100kilometer cycling time trials (22, 35, 44). However, Zajac and colleagues (20) provide new insights revealing that competitive off-road cyclists undergoing 4 weeks of a KD experienced reduced exercising RER concomitant to diminished maximal workload and workload at lactate threshold (45). Collectively, current findings surrounding the KD and aerobic exercise performance, though inconclusive, highlight the need for further examination of how training status impacts adaptation to the KD and resultant performance. Moreover, if the KD does not impair higher intensity aerobic intensity exercise performance (<80%  $\text{VO}_2\text{max}$ ), future evidence needs to support both morphological and functional changes in mitochondria that would allow the cell to meet the increased rate of ATP demand requisite during high intensity aerobic exercise.

#### *Ketogenic Diet and Anaerobic Exercise*

The existing pool of knowledge vis-à-vis the KD and anaerobic performance is scarce. In fact, equivocal results from a mere six studies and one abstract currently encompass the subject. Four studies report that the KD negatively impacts anaerobic performance. Of these, one study utilized recreationally trained subjects and found that KD significantly reduced isotonic strength as measured by a three-set squat repetition total at 80% 1RM (46). Two other studies utilized cycle ergometer to measure anaerobic power output in healthy non-highly trained subjects but their results differed slightly. While one found that KD limited both mean and peak power, the other noted only a decrement in mean power (30, 40). In the former study, the authors noted that the reduction in mean power was mitigated when corrected for a loss in body mass in the KD group. These findings corroborate additional findings in which well-trained cyclists subjected to 6-days of HFD followed by 1-day of CHO loading exhibited reduced exercising fat oxidation rates and impaired high intensity cycle sprint performance (44).

Still, three other studies provide evidence supporting the use of the KD in conjunction with strength and power training; the most compelling of which showed that elite female gymnasts following a KD during normal training did not exhibit impairments in muscular strength, endurance or power (41). Significantly,

this was the only investigation of athletes. Likewise, a pilot study found that KD did not deter gains in either strength or anaerobic power in response to an 8-week periodized resistance-training regimen (47).

One other study highlighted the clinical utility of the KD in showing that overweight women lost body fat while maintaining lean body mass when combining a KD with resistance training (48-50).

Worth noting, two of the aforementioned studies that approximated glycolytic flux while evaluating the effect of KD on anaerobic power and muscular strength generated similar results. Both studies reported depressed blood lactate levels in subjects adhering to a KD immediately following 30-second supra-maximal intensity cycling attempt and 3 sets of squats (30, 51). The authors of each study speculated that this might account for the observed reductions in power and strength since lactate production is positively correlated with glycolytic capacity during high intensity efforts.

The overarching limitation of the above studies is that none allowed for adequate adaptation to the KD, allotting a dubious task of synthesizing interpretations. Future studies must allow for at least 8-weeks for diet adaptation before testing commences. Furthermore, significant attention must be given to how adaptation to the KD alters intramyocellular energetics during anaerobic exercise. Studies must specify creatine phosphate (CP) turnover rates during sets and rest periods of anaerobic exercise as well as whether KD affects intramyocellular CP storage levels. Finally, modalities of anaerobic activities must be diversified, i.e. resistance based training, and should aim to better mimic “real world” anaerobic exercise.

### Practical Application of the KD

Though the precise impact of ketosis on exercise performance remains unclear, practitioners looking to integrate this nutrition strategy into their training may consider several methods. Dietary induction of ketosis is most successful when >60% caloric energy is derived from fat and <5% is from CHO (52). Protein should constitute a significant proportion of dietary energy (>20%) as it retards muscle wasting and amino acids such as leucine and lysine are ketogenic (53). Upon absorption, medium chain triglycerides (MCT's) avoid systemic circulation and instead enter portal circulation for immediate oxidation by the liver, making MCT oil a possible adjuvant to a KD (54). However, tolerability of MCT oil, especially at higher doses, may be individualized, thus care should be taken when considering this method (55). Beyond the scope of nutritional and supplemental methods, is an exciting new patent for a synthetic ketone body and ketone ester, providing an intriguing new possibility for inducing ketosis, preventing muscle glycogen breakdown, aiding muscle recovery and preventing muscle wasting. Before undertaking a KD, the practitioner must consider performance goals, health and nutritional access in an effort to maximize effectiveness, adherence and safety.

*Acknowledgements.* The author declares no conflict of interest. The study was not funded. The results of the present study do not constitute endorsement of the product by the author.

### References

1. Sharman MJ, Kraemer WJ, Love DM, Avery NG, Gómez AL, Scheett TP, Volek JS. (2002). A ketogenic diet favorably affects serum biomarkers for cardiovascular disease in normal-weight men. *J Nutr*; 132(7): 1879-85.
2. Hartman AL, Vining EP (2007). Clinical aspects of the ketogenic diet. *Epilepsia*; 48(1): 31-42.
3. Paoli A, Damiani E, Bosco G (2014). Ketogenic diet in neuromuscular and neurodegenerative diseases. *Biomed Res Int*. ID 474296. <http://dx.doi.org/10.1155/2014/474296>. Paoli, A., et al., *Ketogenic diet in neuromuscular and neurodegenerative diseases*. *Biomed Res Int*, 2014. **2014**: p. 474296.
4. Paoli, A., L. Cenci, and K.A. Grimaldi (2011). Effect of ketogenic Mediterranean diet with phytoextracts and low carbohydrates/high-protein meals on weight, cardiovascular risk factors, body composition and diet compliance in Italian council employees. *Nutr J*, 10: 112.
5. Maalouf M, Rho JM, Mattson MP (2009). The neuroprotective properties of calorie restriction, the ketogenic diet, and ketone bodies. *Brain Res Rev*; 59(2): 293-315.
6. Abdelwahab MG, Fenton KE, Preul MK, Rho JM, Lynch A, Stafford P, Scheck AC (2012). The ketogenic diet is an effective adjuvant to radiation therapy for the treatment of malignant glioma. *PLoS One*; 7(5): p. e36197.
7. Woolf EC, Scheck AC (2015). The ketogenic diet for the treatment of malignant glioma. *J Lipid Res*; 56(1): 5-10.
8. Poff AM, Ari C, Seyfried TN, D'Agostino DP (2013). The ketogenic diet and hyperbaric oxygen therapy prolong survival in mice with systemic metastatic cancer. *PLoS One*; 8(6): p. e65522.
9. Seyfried BT, Kiebish M, Marsh J, Mukherjee P (2009). Targeting energy metabolism in brain cancer through calorie restriction and the ketogenic diet. *J Cancer Res Ther*; 5 Suppl 1: S7-15.

10. Stafford P, Abdelwahab MG, Kim DY, Preul MC, Rho JM, Scheck AC. (2010). The ketogenic diet reverses gene expression patterns and reduces reactive oxygen species levels when used as an adjuvant therapy for glioma. *Nutr Metab (Lond)*, 2010. 7: p. 74.
11. Brownlow ML, Benner L, D'Agostino D, Gordon MN, Morgan D (2013). Ketogenic diet improves motor performance but not cognition in two mouse models of Alzheimer's pathology. *PLoS One*, 2013. 8(9): p. e75713.
12. Cox PJ, Clarke K (2014). Acute nutritional ketosis: implications for exercise performance and metabolism. *Extrem Physiol Med*; 3: p. 17.
13. Bergstrom J, Hermansen L, Hultman E, Saltin B (1967). Diet, muscle glycogen and physical performance. *Acta Physiol Scand*; 71(2): p. 140-50.
14. Thomas DT, Erdman KA, Burke LM (2016). American College of Sports Medicine Joint Position Statement. Nutrition and Athletic Performance. *Med Sci Sports Exerc*; 48(3): 543-68.
15. Volek JS, Sharman MJ, Love DM, Avery NG, Gómez AL, Scheett TP, Kraemer WJ (2002). Body composition and hormonal responses to a carbohydrate-restricted diet. *Metabolism*. 51(7): 864-70.
16. Cotter DG, Schugar RC, Crawford PA (2013). Ketone body metabolism and cardiovascular disease. *Am J Physiol Heart Circ Physiol*; 304(8): H1060-76.
17. Cahill GF Jr (2006). Fuel metabolism in starvation. *Annu Rev Nutr*; 26: p. 1-22.
18. Badman MK (2009). A very low carbohydrate ketogenic diet improves glucose tolerance in ob/ob mice independently of weight loss. *Am J Physiol Endocrinol Metab*; 297(5): p. E1197-204.
19. Woolf EC, Curley KL, Liu Q, Turner GH, Charlton JA, Preul MC, Scheck AC (2015). The Ketogenic Diet Alters the Hypoxic Response and Affects Expression of Proteins Associated with Angiogenesis, Invasive Potential and Vascular Permeability in a Mouse Glioma Model. *PLoS One*; 10(6): p. e0130357.
20. Miller WC, Bryce GR, Conlee RK (1984). Adaptations to a high-fat diet that increase exercise endurance in male rats. *J Appl Physiol Respir Environ Exerc Physiol*; 56(1): p. 78-83.
21. Simi B, Sempore B, Mayet MH, Favier RJ (1985). Additive effects of training and high-fat diet on energy metabolism during exercise. *J Appl Physiol*; 71(1):197-203.
22. Goedecke JH, Christie C, Wilson G, Dennis SC, Noakes TD, Hopkins WG, Lambert EV (1999) . Metabolic adaptations to a high-fat diet in endurance cyclists. *Metabolism*; 48(12): 1509-17.
23. Helge JW, Kiens B (1997). Muscle enzyme activity in humans: role of substrate availability and training. *Am J Physiol*; 272(5 Pt 2): p. R1620-4.
24. Cheng B, Karamizrak O, Noakes TD, Dennis SC, Lambert EV (1997). Time course of the effects of a high-fat diet and voluntary exercise on muscle enzyme activity in Long-Evans rats. *Physiol Behav*; 61(5): p. 701-5.
25. Kennedy AR, Pissios P, Otu H, Roberson R, Xue B, Asakura K, Furukawa N, Marino FE, Liu FF, Kahn BB, Libermann TA, Maratos-Flier E (2007). A high-fat, ketogenic diet induces a unique metabolic state in mice. *Am J Physiol Endocrinol Metab*; 292(6): p. E1724-39.
26. Cutler DL, Gray CG, Park SW, Hickman MG, Bell JM, Kolterman OG (1995). Low-carbohydrate diet alters intracellular glucose metabolism but not overall glucose disposal in exercise-trained subjects. *Metabolism*; 44(10): 1264-70.
27. Rosholt MN, King PA, Horton ES (1994). High-fat diet reduces glucose transporter responses to both insulin and exercise. *Am J Physiol*; 266(1 Pt 2): p. R95-101.
28. Achten J, Jeukendrup AE (2003). Maximal fat oxidation during exercise in trained men. *Int J Sports Med*; 24(8): 603-8.
29. Lambert EV, Speechly DP, Dennis SC, Noakes TD (1994). Enhanced endurance in trained cyclists during moderate intensity exercise following 2 weeks adaptation to a high fat diet. *Eur J Appl Physiol Occup Physiol*; 69(4): 287-93.
30. Langfort J, Zarzeczny R, Pilis W, Nazar K, Kaciuba-Uściłko H (1997). The effect of a low-carbohydrate diet on performance, hormonal and metabolic responses to a 30-s bout of supramaximal exercise. *Eur J Appl Physiol Occup Physiol*; 76(2): 128-33.
31. Phinney SD, Horton ES, Sims EA, Hanson JS, Danforth E Jr, LaGrange BM (1980). Capacity for moderate exercise in obese subjects after adaptation to a hypocaloric, ketogenic diet. *J Clin Invest*; 66(5): 1152-61.
32. Volek J, Freidenreich DJ, Saenz C, Kunces LJ, Creighton BC, Bartley JM, et al. (2015). Metabolic characteristics of keto-adapted ultra-endurance runners. *Metabolism*. 65(3): 100-110.
33. Hyypää S, Saastamoinen M, Reeta Poso A (1999). Effect of a post exercise fat-supplemented diet on muscle glycogen repletion. *Equine Vet J Suppl*; (30): 493-8.
34. Fournier PA, Fairchild TJ, Ferreira LD, Bräu L (2004). Post-exercise muscle glycogen repletion in the extreme: effect of food absence and active recovery. *J Sports Sci Med*; 3(3): 139-46.
35. Lambert EV, Goedecke JH, Zyle C, Murphy K, Hawley JA, Dennis SC, Noakes TD (2001). High-fat diet versus habitual diet prior to carbohydrate loading: effects of exercise metabolism and cycling performance. *Int J Sport Nutr Exerc Metab*; 11(2): 209-25.
36. Phinney SD, Bistrian BR, Evans WJ, Gervino E, Blackburn GL (1983). The human metabolic response to chronic ketosis without caloric restriction: preservation of submaximal exercise capability with reduced carbohydrate oxidation. *Metabolism*; 32(8): 769-76.

37. Benoit FL, Martin RL, Watten RH (1965). Changes in body composition during weight reduction in obesity. Balance studies comparing effects of fasting and a ketogenic diet. *Ann Intern Med*; 63(4): 604-12.
38. Westman EC, Yancy WS, Edman JS, Tomlin KF, Perkins CE (2002). Effect of 6-month adherence to a very low carbohydrate diet program. *Am J Med*; 113(1): 30-6.
39. Willi SM, Oexmann MJ, Wright NM, Collop NA, Key LL Jr (1998). The effects of a high-protein, low-fat, ketogenic diet on adolescents with morbid obesity: body composition, blood chemistries, and sleep abnormalities. *Pediatrics*; 101(1 Pt 1): p. 61-7.
40. Fleming J, Sharman MJ, Avery NG, Love DM, Gómez AL, Scheett TP, Kraemer WJ, Volek JS (2003). Endurance capacity and high-intensity exercise performance responses to a high fat diet. *Int J Sport Nutr Exerc Metab*; 13(4): p. 466-78.
41. Paoli A, Grimaldi K, D'Agostino D, Cenci L, Moro T, Bianco A, Palma A (2012). Ketogenic diet does not affect strength performance in elite artistic gymnasts. *J Int Soc Sports Nutr*; 9(1): p. 34.
42. Claudia M. Meirelles, P.S.C.G. (2016). Effects Of Short-Term Carbohydrate Restrictive And Conventional Hypoenergetic Diets And Resistance Training On Strength Gains And Muscle Thickness. *Journal of Sport Science and Medicine*; 15: 578-584.
43. Rhyu HS, Cho SY (2014). The effect of weight loss by ketogenic diet on the body composition, performance-related physical fitness factors and cytokines of Taekwondo athletes. *J Exerc Rehabil*; 10(5): 326-31.
44. Havemann L, West SJ, Goedecke JH, Macdonald IA, St Clair Gibson A, Noakes TD, Lambert EV (2006). Fat adaptation followed by carbohydrate loading compromises high-intensity sprint performance. *J Appl Physiol* (1985); 100(1): 194-202.
45. Zajac A, Poprzecki S, Maszczyk A, Czuba M, Michalczyk M, Zydek G (2014). The effects of a ketogenic diet on exercise metabolism and physical performance in off-road cyclists. *Nutrients*; 6(7): 2493-508.
46. Leveritt M, Abernethy PJ (1999). Effects of Caloric Restriction on Strength Performance. *Journal of Strength and Conditioning Research*; 13(1): p. 52-57.
47. McCleary SA, Sharp MH, Lowery RP, Silva JE, Rauch JT, Ormes JA, Shields KA, Georges JI, Wilson JM (2014). Effects of a ketogenic diet on strength and power, in The Eleventh International Society of Sports Nutrition (ISSN) Conference and Expo 2014: Clearwater Beach, FL.
48. Jabekk PT, Moe IA, Meen HD, Tomten SE, Høstmark AT (2010). Resistance training in overweight women on a ketogenic diet conserved lean body mass while reducing body fat. *Nutr Metab (Lond)*; 7: 17.
49. Leveritt M, Abernethy PJ, Barry BK, Logan PA (1999). Concurrent strength and endurance training. A review. *Sports Med*; 28(6): p. 413-27.
50. Apostol Adela, Ionescu AM, Vasilescu M (2013). Aerobic versus Anaerobic - comparative studies concerning the dynamics of the aerobic and anaerobic effort parameters in top athletes. *Medicina Sportiva*; IX(2): 2130-2140.
51. Volek J, Sharman MJ, Gómez AL, Judelson DA, Rubin MR, Watson G et al. (2004). Comparison of energy-restricted very low-carbohydrate and low-fat diets on weight loss and body composition in overweight men and women. *Nutr Metab (Lond)*; 1(1): 13.
52. Berg J, Tymoczko J, Stryer L (2002). *Amino Acid Catabolism*, in *Biochemistry*, L. Stryer, Editor. 2002, WH Freeman: New York.
53. Liu YM, Wang HS (2013). Medium-chain triglyceride ketogenic diet, an effective treatment for drug-resistant epilepsy and a comparison with other ketogenic diets. *Biomed J*; 36(1): p. 9-15.
54. Azzam R, Azar NJ (2013). Marked Seizure Reduction after MCT Supplementation. *Case Rep Neurol Med*; 809151.
55. Clarke K, Cox P (2015). *Ketone body and ketone body ester for reducing muscle breakdown* U.S.P.a.T. Office, Editor. TDELTA LIMITED, Thame (GB): United States.

*Corresponding author*

Evan E. Schick  
Department of Kinesiology, HHS2-210  
California State University, Long Beach  
1250 Bellflower Ave.  
Long Beach, CA 90840  
Email: [evan.schick@csulb.edu](mailto:evan.schick@csulb.edu)  
Tel: (562) 985-4590

Received: September 6, 2016

Accepted: November 25, 2016